

## Withanolides from *Hyoscyamus niger* Seeds

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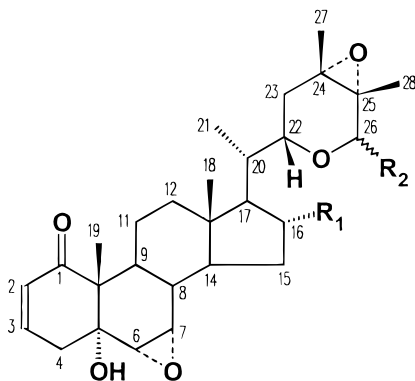
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Three withanolide class steroids were isolated from the seeds of *Hyoscyamus niger*. Two of them were identified as daturalactone-4 (**1**) and Nic-3 (which is now named hyoscyamilactol) (**2**). The new compound was elucidated as 16 $\alpha$ -acetoxyhyoscyamilactol (**3**) on the basis of spectroscopic properties and X-ray crystallographic analysis.

The genus *Hyoscyamus* (Solanaceae) is well-known for the production of anticholinergic tropane alkaloids such as hyoscyamine and scopolamine (hyoscine). In Chinese medicine, the seeds of *H. niger* (known as *Tian-Xian-Zi*, literally meaning Fairy Lady Seeds) are used as an antispasmodic, sedative and analgesic agent, prescribed for the treatment of stomach cramps, heavy coughs, neuralgia, and manic psychosis.<sup>1</sup> As part of a continuing project to study the chemical composition of toxic Chinese herbal drugs, we have examined the seeds of *H. niger*. Analytical methods have been established for the major alkaloids by using HPLC<sup>2</sup> and capillary electrophoresis (unpublished results).

This paper deals with the isolation and structural determination of three withanolide derivatives. An EtOH extract of the seeds of *H. niger* was concentrated and partitioned between CHCl<sub>3</sub> and H<sub>2</sub>O. The CHCl<sub>3</sub> fraction afforded three steroidal compounds (**1**–**3**) upon repeated chromatography on silica gel.



- (1) R<sub>1</sub>=H; R<sub>2</sub>=O  
 (2) R<sub>1</sub>=H; R<sub>2</sub>= $\alpha$ -OH,  $\beta$ -H  
 (3) R<sub>1</sub>=OAc; R<sub>2</sub>= $\alpha$ -OH,  $\beta$ -H

Daturalactone-4 (**1**) was obtained as colorless needles. The molecular formula C<sub>28</sub>H<sub>38</sub>O<sub>6</sub> was derived from the DCI-MS ([M + 1]<sup>+</sup> at *m/z* 471) and <sup>13</sup>C NMR data. Its spectral properties were identical in all respects (IR, NMR, MS) with those of daturalactone-4, a withanolide steroid previously isolated from *Datura quercifolia*.<sup>3</sup>

Compound **2** was obtained as colorless needles. The DCI-MS displayed a quasimolecular ion signal at *m/z* 473 [M + 1]<sup>+</sup> and a base peak at *m/z* 455 [M – OH]<sup>+</sup>. The molecular formula C<sub>28</sub>H<sub>40</sub>O<sub>6</sub> was derived on the basis of the MS result and <sup>13</sup>C NMR data. The IR spectrum exhibited absorption bands of hydroxyl and  $\alpha,\beta$ -unsaturated ketone functional groups, but no absorption was observed for the lactone carbonyl. The <sup>1</sup>H and <sup>13</sup>C NMR spectra clearly suggested that **2** was a lactol derivative of **1**. In the literature, a structure identical to that of **2** (coded as Nic-3<sup>4</sup>) was described from the leaves of *Nicandra physaloides*, possessing inhibitory effect on the feeding of insect larvae.<sup>4</sup> More recently the same compound was found in the whole plant of *Exodeconus maritimus*.<sup>5</sup> The X-ray structure of its 26-O-acetate derivative was reported.<sup>4</sup> We now propose a trivial name of hyoscyamilactol for **2**, and provide NMR assignments based on <sup>1</sup>H–<sup>1</sup>H COSY, HETCOR, and COLOC results.

Compound **3** was obtained as colorless crystals. The DCI-MS contained a quasimolecular ion signal at *m/z* 531 [M + 1]<sup>+</sup>, a base peak at *m/z* 513 [M – OH]<sup>+</sup>, and a prominent fragment peak at *m/z* 471 [M – OAc]<sup>+</sup>. The molecular formula C<sub>30</sub>H<sub>42</sub>O<sub>8</sub> was derived from the MS and <sup>13</sup>C NMR data. Examination of the NMR spectra led to the identification of a lactol ring ( $\delta_C$  91.8 and  $\delta_H$  4.97) and an acetoxy group ( $\delta_C$  170.9, 21.2 and  $\delta_H$  2.08). Compound **3** was therefore an acetoxy derivative of **2**, and attention was then focused on the location of the acetoxy group. Examination of the HMBC spectrum revealed that both <sup>13</sup>C signals at  $\delta$  37.7 (C-20) and 49.1 (C-14) displayed long-range couplings with the methine group ( $\delta_C$  78.6,  $\delta_H$  5.00) bearing the acetoxy substituent. The <sup>1</sup>H–<sup>1</sup>H COSY spectrum also showed couplings between the signal at  $\delta$  5.00 and those at  $\delta$  1.41 (H-17) and 1.92 (H-15). On the basis of these findings, it was concluded that the acetoxy substituent was located on C-16 of the steroid structure, and compound **3** was the 16-acetoxy derivative of **2**, i.e., 16-acetoxyhyoscyamilactol.

Finally, a suitable single crystal of **3** was chosen for X-ray crystallographic analysis to confirm the planar structure as well as its relative stereochemistry. The crystal structure is shown in Figure 1, and the bond lengths for **3** are given in Table 1. With the assumption that all other chiral centers conformed to the stereochemistry of daturalactone-4 (**1**), the lactol hydroxyl group in **2** and **3** was  $\alpha$ -oriented and thus in a syn relationship with the epoxy ring. The 16-acetoxy functional group in **3** was determined to be in an  $\alpha$  orientation; and **3** is therefore 16 $\alpha$ -acetoxyhyoscyamilactol. Although withanolides commonly occur in Solanaceae plants<sup>6</sup> and 16-hydroxylated derivatives are

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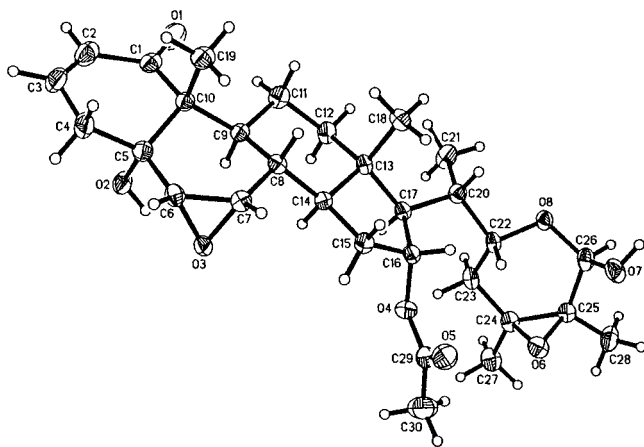


Figure 1. Crystal structure of **3**.

Table 1. Bond Lengths (Å) of **3**

O(1)–C(1)	1.214 (8)	O(2)–C(5)	1.444 (9)
O(3)–C(6)	1.436 (5)	O(3)–C(7)	1.448 (7)
O(4)–C(16)	1.443 (6)	O(4)–C(29)	1.339 (7)
O(5)–C(29)	1.212 (10)	O(6)–C(24)	1.453 (8)
O(6)–C(25)	1.443 (8)	O(7)–C(26)	1.407 (9)
O(8)–C(22)	1.456 (7)	O(8)–C(26)	1.419 (5)
C(1)–C(2)	1.486 (9)	C(1)–C(10)	1.522 (7)
C(2)–C(3)	1.315 (11)	C(3)–C(4)	1.478 (9)
C(4)–C(5)	1.517 (7)	C(5)–C(6)	1.517 (7)
C(5)–C(10)	1.550 (9)	C(6)–C(7)	1.460 (7)
C(7)–C(8)	1.498 (9)	C(8)–C(9)	1.542 (8)
C(8)–C(14)	1.525 (6)	C(9)–C(10)	1.560 (7)
C(9)–C(11)	1.534 (9)	C(10)–C(19)	1.543 (9)
C(11)–C(12)	1.545 (6)	C(12)–C(13)	1.530 (8)
C(13)–C(14)	1.545 (9)	C(13)–C(17)	1.548 (5)
C(13)–C(18)	1.547 (9)	C(14)–C(15)	1.520 (7)
C(15)–C(16)	1.534 (7)	C(16)–C(17)	1.536 (8)
C(17)–C(20)	1.554 (8)	C(20)–C(21)	1.536 (9)
C(20)–C(22)	1.539 (5)	C(22)–C(23)	1.510 (9)
C(23)–C(24)	1.504 (5)	C(24)–C(25)	1.462 (9)
C(24)–C(27)	1.498 (9)	C(25)–C(26)	1.513 (8)
C(25)–C(28)	1.506 (6)	C(29)–C(30)	1.469 (9)

known,<sup>5</sup> 16 $\alpha$ -acetoxyhyoscyamilactol is hitherto unreported in the literature.

## Experimental Section

**General Experimental Procedures.** The melting points were measured on a Leica Galen III apparatus and were uncorrected. Optical rotation was measured using a Perkin-Elmer 241 polarimeter. IR and UV spectra were recorded on a Perkin-Elmer 16PC FTIR spectrometer and a Milton Roy Spectronic 3000 array spectrophotometer, respectively. <sup>1</sup>H and <sup>13</sup>C NMR data were obtained from a JEOL JNM-EX-400 FT-NMR spectrometer at 400 and 98.5 MHz, respectively, with TMS as internal standard. DCI-MS (CH<sub>4</sub>) were determined on a Finnigan TSQ7000 triple quadrupole mass spectrometer. The X-ray crystallographic data were obtained using a Siemens P4/RA diffractometer.

**Plant Materials.** The seeds of *H. niger* were obtained from an herbal drug supplier in Beijing in 1994 and authenticated at the National Institute for the Control of Pharmaceutical and Biological Products, Beijing. A voucher specimen has been deposited in the herbarium of the same institution.

**Extraction and Isolation.** Air-dried seeds of *H. niger* (5 kg) were ground and defatted with hexane before being extracted with 95% EtOH at room temperature. The EtOH extract was concentrated under reduced pressure and partitioned between hexane and H<sub>2</sub>O. The aqueous portion was washed with CHCl<sub>3</sub>, EtOAc, and *n*-BuOH successively. A residue (2.5 g) was obtained from the CHCl<sub>3</sub> fraction after evaporation of excessive solvent under reduced pressure. This residue was chromatographed over silica gel using mixtures

of CHCl<sub>3</sub>–acetone of increasing polarities. The effluents were combined into 10 fractions on the basis of TLC patterns. Fractions 3 and 6 were further purified by silica gel chromatography washed with CHCl<sub>3</sub>–acetone mixtures to afford **1** (30 mg) and **2** (40 mg), respectively. Compound **3** (13 mg) was purified from fraction 8 by crystallization.

**Daturalactone-4 (1):** colorless needles, mp 288–290 °C (lit.<sup>3</sup> 282 °C); [ $\alpha$ ]<sub>D</sub> 132.8° (c 0.013, MeOH); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  203.2 (s, C-1), 170.0 (s, C-26), 139.6 (d, C-3), 129.0 (d, C-2), 76.3 (d, C-22), 73.2 (s, C-5), 62.2 (s, C-25), 59.3 (s, C-24), 57.2 (d, C-7), 56.3 (d, C-6), 51.9 (d, C-17), 51.3 (d, C-14), 51.0 (s, C-10), 43.5 (s, C-13), 39.8 (t, C-12), 38.7 (d, C-20), 36.7 (t, C-4), 35.7 (d, C-8), 35.5 (d, C-9), 28.7 (t, C-23), 27.2 (t, C-16), 23.5 (t, C-15), 21.8 (t, C-11), 17.9 (q, 27-CH<sub>3</sub>), 14.6 (q, 19-CH<sub>3</sub>), 13.6 (q, 28-CH<sub>3</sub>), 12.9 (q, 21-CH<sub>3</sub>), 12.0 (q, 18-CH<sub>3</sub>); DCI-MS (CH<sub>4</sub>)  $m/z$  471 (100%) [M + 1]<sup>+</sup>.

**Hyoscyamilactol (2):** colorless needles, mp 241–243 °C (lit.<sup>4</sup> 248 °C); [ $\alpha$ ]<sub>D</sub> 98.4° (c 0.012, MeOH); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  203.2 (s, C-1), 139.5 (d, C-3), 129.0 (d, C-2), 91.7 (d, C-26), 73.2 (s, C-5), 65.2 (d, C-22), 65.0 (s, C-25), 63.8 (s, C-24), 57.3 (d, C-7), 56.3 (d, C-6), 52.4 (d, C-17), 51.4 (d, C-14), 51.0 (s, C-10), 43.4 (s, C-13), 39.9 (t, C-12), 39.1 (d, C-20), 36.7 (t, C-4), 35.7 (d, C-8), 35.5 (d, C-9), 29.4 (t, C-23), 27.2 (t, C-16), 23.5 (t, C-15), 21.9 (t, C-11), 18.9 (q, 28-CH<sub>3</sub>), 16.5 (q, 27-CH<sub>3</sub>), 14.7 (q, 19-CH<sub>3</sub>), 12.6 (q, 21-CH<sub>3</sub>), 12.1 (q, 18-CH<sub>3</sub>); DCI-MS (CH<sub>4</sub>)  $m/z$  473 [M + 1]<sup>+</sup>, 455 (100%) [M – OH]<sup>+</sup>.

**16 $\alpha$ -Acetoxyhyoscyamilactol (3):** colorless plates, mp 262–264 °C; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.60 (1H, ddd,  $J$  = 10, 5, 2 Hz, H-3), 5.85 (1H, dd,  $J$  = 10, 2.5 Hz, H-2), 5.00 (1H, m, H-16), 4.97 (1H, s, br s, H-26), 3.54 (1H, dt,  $J$  = 3.0, 11.5 Hz, H-22), 3.49 (1H, br s, D<sub>2</sub>O exchangeable, 26-OH), 3.25 (1H, br d,  $J$  = 4 Hz, H-7), 3.14 (1H, s, D<sub>2</sub>O exchangeable, 5-OH), 3.05 (1H, d,  $J$  = 4 Hz, H-6), 2.68 (1H, br d,  $J$  = 19 Hz, H-4a), 2.53 (1H, dd,  $J$  = 19, 5 Hz, H-4b), 2.08 (3H, s, COCH<sub>3</sub>), 1.41 (3H, s, 27-CH<sub>3</sub>), 1.32 (3H, s, 28-CH<sub>3</sub>), 1.17 (3H, s, 19-CH<sub>3</sub>), 0.93 (3H, d,  $J$  = 7 Hz, 21-CH<sub>3</sub>), 0.79 (3H, s, 18-CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  203.1 (s, C-1), 170.9 (s, COCH<sub>3</sub>), 139.6 (d, C-3), 129.0 (d, C-2), 91.8 (d, C-26), 78.6 (d, C-16), 73.2 (s, C-5), 65.0 (d, C-22), 65.0 (s, C-25), 63.8 (s, C-24), 57.9 (d, C-17), 57.0 (d, C-7), 56.2 (d, C-6), 50.9 (s, C-10), 49.1 (d, C-14), 44.1 (s, C-13), 39.8 (t, C-12), 37.7 (d, C-20), 36.7 (t, C-4), 35.5 (d, C-9), 35.2 (d, C-8), 33.6 (t, C-15), 29.5 (t, C-23), 21.5 (t, C-11), 21.2 (q, COCH<sub>3</sub>), 18.9 (q, 28-CH<sub>3</sub>), 16.5 (q, 27-CH<sub>3</sub>), 14.7 (q, 19-CH<sub>3</sub>), 13.4 (q, 18-CH<sub>3</sub>), 12.8 (q, 21-CH<sub>3</sub>); DCI-MS (CH<sub>4</sub>)  $m/z$  531 [M + 1]<sup>+</sup>, 513 (100%) [M – OH]<sup>+</sup>, 471 [M – CH<sub>3</sub>COO]<sup>+</sup>.

**Crystal Data for 3:** C<sub>30</sub>H<sub>42</sub>O<sub>8</sub>,  $M$  = 530.6, colorless plates, 2.0  $\times$  0.4  $\times$  0.05 mm, monoclinic, space group  $C_2$ ,  $T$  = 298 K,  $a$  = 26.065(5) Å,  $b$  = 5.948(2) Å,  $c$  = 21.075(5) Å,  $\beta$  = 123.41(2)°,  $V$  = 2727.1(12) Å<sup>3</sup>,  $Z$  = 4,  $D$  = 1.292 mg/m<sup>3</sup>,  $F(100)$  = 1144,  $\mu$ (Mo K $\alpha$ ) = 0.093 mm<sup>-1</sup>,  $a$  = 0.710 37 Å.

**Data Collection and Structure Refinement.** Intensities of 3099 reflections were collected on a Siemens P4-RA diffractometer operating at 1 kW using the  $\omega/2\theta$  scan mode up to  $2\theta$  = 53°. Of these, 2233 were unique and observed  $F \geq 4\sigma(F)$ . The structure was solved by direct methods and refined (343 parameters) by full-matrix least-squares methods using the SHELXTL suite of X-ray programs.<sup>8</sup> All non-hydrogen atoms were refined using anisotropic thermal parameters. Hydrogen atoms were located in difference Fourier maps and then positioned using geometric constraints. A common isotropic thermal parameter was refined for all hydrogens. The final refinement converged with  $R$  = 5.79%,  $wR$  = 5.93%, and goodness-of-fit = 1.74. The largest difference peak and largest difference hole in the final difference Fourier map were 0.31 and –0.29 e Å<sup>-3</sup>, respectively. The absolute configuration was not determined.

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## References and Notes

- 1) *Chinese Pharmacopoeia*, 1995 ed.; Vol. 1, p. 41.

- (2) Ma, C. Y.; Chen, D. C.; Yang, Z. Q. *Chin. J. Pharm. Anal.* **1996**, *16*, 119–121.
- (3) Qurishi, M. A.; Kalla, A. K.; Dhar, K. L. *Phytochemistry* **1979**, *18*, 1756–1757.
- (4) Begley, M. J.; Crombie, L.; Ham, P. J. *J. Chem. Soc., Perkin Trans.* **1976**, 296–304.
- (5) Gil, R. R.; Misico, R. I.; Sotes, I. R.; Oberti, J. C. *J. Nat. Prod.* **1997**, *60*, 568–572.
- (6) Ray, A. B.; Gupta, M. in *Progress in the Chemistry of Organic Natural Products*, Springer-Verlag: New York, 1994; Vol. 63, pp 1–106.
- (7) Crystallographic data for the structure reported in this paper have been deposited with the Cambridge Crystallographic Data Centre. Copies of the data can be obtained, free of charge, on application to the Director, CCDC, 12 Union Road, Cambridge CB2 1EZ, UK (fax, +44 (0)1223 336033 or e-mail, deposit@ccdc.cam.ac.uk).
- (8) Sheldrick, G. M. *SHELXTL*; Siemens Analytical Instruments: Madison, WI, 1993.

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